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THE SYNTHESIS OF FLUORINATED AMINOPHOSPHONIC ACID INHIBITORS OF ALANINE RACEMASE Gary A. Flynn*, Douglas W. Beight, Ekkehard H.W. Bohme, and Brian W. Metcalf

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ABSTRACT: The synthesis of β -trifluoro, β -difluoro, and β -monofluoro-1-aminoethanephosphonic acids is described utilizing fluorinated acetic acids as starting materials.

The inhibition of peptidoglycan biosynthesis continues to be a primary focus of antibacterial drug design. The intrinsic property of bacteria to incorporate D-alanine into their cell wall has prompted recent efforts to inhibit the essential pyridoxal dependent racemase responsible for the production of D-alanine. Cycloserine^{1,2} O-carbamoylserine^{2,3} and the β -haloalanines^{2,4} <u>1</u> are mechanism-based inactivators of bacterial alanine racemase whose presumed mechanism of action⁵ is illustrated in Scheme I. In contrast, L-1-aminoethanephosphonic acid <u>2</u> is a potent selective competitive inhibitor of alanine racemase <u>in vitro</u> which when converted to the L-alanine dipeptide <u>5</u> (Alaphosphin) displays significant antibacterial properties and synergism with D-cycloserine and β -lactam antibiotics.^{6,7}



SCHEME I

The fluorinated aminophosphonic acids $\underline{3a-c}$ were considered to be attractive potential inhibitors of alanine racemase which might exhibit the enhanced binding affinity and specificity of aminophosphonic acid $\underline{2}$ while retaining the irreversible vector of the fluorinated alanines.^{2,4}



Our general synthetic approach to all three fluorinated aminophosphonic acids <u>3a-c</u> (Scheme II) would, in principle, utilize the corresponding trifluoro, difluoro, and monofluoroacetic acids <u>4a-c</u> as starting materials. Due to the extreme toxicity of fluoroacetic acid <u>4c</u> $(LD_{50}=7mg/kg)$, our exploratory efforts began with the condensation of trifluoroacetic anhydride with 1-aminodiphenylmethane in pyridine (90% yield). The resulting amide <u>6a</u> was converted to imidoyl chloride <u>7a</u> (1 eq. PCI₅, PhH, reflux, 18 h) in 62% yield⁸ after Kugelrohr distillation (125°C at 2 mm Hg). Heating of equivalent amounts of imidoyl chloride, diethyl phosphite, and triethylamine in the absence of solvent at 60°C for 20 hours gave a 30-40% yield of isomerized imine <u>8</u> after chromatography (Rf=0.20, 20% EtOAc/hexane).⁹ Assignment of the isomerized structure <u>8</u> to this imine was supported by downfield aromatic protons in the NMR.

0 II RCOH +	H ₂ NCHPh ₂	EEDQ	O II RCNHCHF	Рh ₂ РСI3. Рhн Д	>
4ª R=CF₃ 4b R=CHF₂ ↔	5		60 60 60		
CI RC=NCHPh ₂ 7_0 7_b \downarrow (E10) _s P:	о (Е (ЕЮ) ₂ ён (Е Еі _з №÷	(10) ₂ P=0 CF ₃ CH−N 8	I=CPh ₂	1) Hg/Pd/C 2)HCI	90 ~
$(E + 0)_2 P = 0$ $R - C = N - CHPt$ $IOa \qquad IOb$ $\int DBU$	NoBH ₃ CN HOAC	(EtO) ₂ P= R – CH		H ₂ Pd/C	ያ እ እ
(E10)₂P=0 H ← N≈CPh F	NoBH ₃ CN HOAc→ (R'(R	(Et0) ₂ P= FCH ₂ CH)) ₂ P=0 -CH-NH [®] ₃	:0 −NHCHPh₂ !!⊂ ,CI [°]	H _E Pd/C	9¢ ∼
	90 R'=Et, R=CF3 96 R'=Et, R=CHF2 90 R'=Et, R=CH2F		30 R'=H, R=CF3 30 R'=H, R=CHF2 30 R'=H, R=CH2F		

SCHEME II

Hydrogenation of <u>8</u> followed by acidification with HCl gave the analytically pure hydrochloride salt <u>9a</u> as a sublimable white solid, mp. 117-118°C in 30% yield. The poor yield reported for this last step reflects partial phosphonic ester hydrolysis. Direct hydrolysis of the crude hydrogenation product without intermediate isolation (6N HCl, reflux, 6 h) provided the targeted aminophosphonic acid hydrochloride salt 3a in 84% yield.

Application of this initial route to the synthesis of the difluoro analog <u>3b</u> proceeded smoothly to the imidoyl chloride stage. Difluoroacetic acid <u>4b</u> was coupled to amine <u>5</u> (EEDQ, CH_2Cl_2 , 95%) to give amide <u>6b</u> which was quantitatively converted to the thermally unstable imidoyl chloride <u>7b</u>. Treatment of <u>7b</u> with diethyl phosphite under a variety of basic conditions failed to yield any tractable product. We surmised that products from this reaction might be unstable to the basic conditions employed and that loss of HF and subsequent polymerization had occurred. Imidoyl chloride <u>7b</u> was successfully condensed with triethyl phosphite under the more neutral Arbuzov conditions [1 eq. (Et0)₃P, neat, 80°C, 18 h] to give the nonisomerized iminophosphonate <u>10b</u> in 45-50% yield after chromatography (Rf=0.35, 20% Et0Ac/hexane).

In contrast to isomerized imine $\underline{8}$, the relatively hindered imine $\underline{10b}$ was resistant to catalytic hydrogenation and was converted to complex mixtures with severe material loss under forcing conditions. Enamino imine $\underline{12}$ was an interesting by-product isolated in small amounts from several of these attempts. Elimination of HF in this system is apparently a major pathway to decomposition.

The analogous Arbuzov reaction of trifluoroimidoyl chloride <u>7a</u> with triethyl phosphite afforded iminophosphonate <u>10a</u> (56% yield, Rf=0.35, 20% EtOAc/hexane) which was clearly different from imine 8 previously obtained.

Trifluoroimine <u>10a</u> and difluoroimine <u>10b</u> underwent smooth reduction with sodium cyanoborohydride in acetic acid to give benzhydrylamines <u>11a</u> and <u>11b</u> in 95% and 100% yields, respectively. These benzhydrylaminophosphonic esters were hydrogenated (H_2 , 10% Pd/C, EtOH, HCI) and hydrolyzed (6N HCI, reflux, 6 h) to give the corresponding aminophosphonic acid hydrochloride salts <u>3a</u> and <u>3b</u> in excellent overall yield. Compound <u>3a</u> obtained from the Arbuzov sequence was identical to that obtained from our initial approach.

Our efforts to prepare the monofluoro analog $\underline{3c}$ by simple modification of either of the methods described thus far were unsuccessful presumably due to the reactivity of the fluoroacetate moiety. These results and the toxicity of fluoroacetic acid and its derivatives led us to look for alternate methods of generating monofluorinated molecules.

The key observation that enamino imine <u>12</u> was formed as a by-product during the attempted catalytic hydrogenation of iminophosphonate <u>10b</u> prompted us to examine the effects of various dehydrohalogenation conditions on <u>10b</u> in hopes of developing by-product <u>12</u> into an intermediate to the monofluoro analog <u>3c</u>. We generally observed that conditions basic enough to promote elimination also led to unacceptable levels of further decomposition. Even lithium diisopropyl amide at -78°C gave complex mixtures in low yields. Clean conversion of <u>10b</u> to <u>12</u> was ultimately achieved with DBU¹⁰ (THF, 25°C, 5H). The sensitive enamino imine <u>12</u> could be isolated in 55-65% yield after chromatography (Rf=0.40, 1:1 EtOAc/hexane) provided that the

temperatures during workup did not exceed 35°C. Treatment of <u>12</u> with NaBH₃CN in acetic acid affected quantitative reduction of both the enamine and imine moieties to give <u>11c</u>. Hydrogenolysis of <u>11c</u> (H₂, Pd/C) followed by ester hydrolysis (6N HCl, reflux, 6 h) provided β -fluoro-1-aminoethanephosphonic acid <u>3c</u> in 90% overall yield.

All three fluorinated aminoethanephosphonic acids have been prepared in good yield using related synthetic techniques. The unusually complex ${}^{1}H$, ${}^{19}F$, and ${}^{31}P$ spectra of these compounds are consistent with their structures. The preparation of the monofluoro analog avoids the use of potentially toxic fluoroacetate derivatives. All three compounds are time-dependent inactivators of alanine racemase 11 with the monofluoro analog being the most potent. A detailed account will be discussed elsewhere.

REFERENCES

- M. Martinez-Carrion and W.T. Jenkins, <u>J. Biol. Chem.</u>, <u>240</u>, 3547 (1965); R. Rando, <u>Accts.</u> <u>Chem. Res.</u>, <u>8</u>, 281 (1975).
- 2. E. Wang and C. Walsh, Biochemistry, 17, 1313 (1978).
- 3. M.P. Lambert and F.C. Neuhaus, J. Bacteriol., 110, 978 (1972).
- T.S. Soper, W.M. Jones, B. Lerner, M. Trop, and J.M. Manning, <u>J. Biol. Chem.</u>, <u>252</u>, 3170 (1977); J.M. Manning and T.S. Soper, "Enzyme-Activated Irreversible Inhibitors", N. Seiler, M.J. Jung, and J. Koch-Weser, eds. Elsevier/North Holland, New York, 163-176; J. Kollonitsch, L. Barash, F. Kahan, and H. Kropp, <u>Nature</u>, <u>243</u>, 346 (1973); J. Kollonitsch and L. Barash, <u>J. Amer. Chem. Soc.</u>, <u>98</u>, 5591 (1976); E. Wang and C. Walsh, <u>Biochemistry</u>, 20, 7539 (1980).
- J.J. Likos, H. Ueno, R.W. Feldhaus, and D.E. Metzler, <u>Biochemistry</u>, <u>21</u>, 4377-4386 (1982);
 H. Ueno, J.L. Likos, and D.E. Metzler, <u>ibid</u>, <u>21</u>, 4387-4393 (1982); B. Badet, K. Lee, H.G. Floss, and C.T. Walsh, <u>Chemm. Commun.</u>, 838-840 (1984).
- J.G. Allen, F.R. Atherton, M.J. Hall, C.H. Hassall, S.W. Holmes, R.W. Lambert, L.J. Nisbet, and R.S. Ringrose, <u>Antimicrob. Agents and Chemo.</u>, <u>15</u>, 684 (1979).
- F.R. Atherton, M.J. Hall, C.H. Hassall, R.W. Lambert, and P.S. Ringrose, <u>ibid</u>, <u>15</u>, 677 (1979).
- 8. Crude yields were nearly quantitative.
- 9. Addition of dialkyl phosphites to imines has been reported: J. Paulsen and H. Kuhne, Chem. Ber., 108, 1239 (1975).
- 10. Diazabicyclo[5.4.0]undec-7-ene (DBU) is available from Aldrich Chemical Company.
- Enzyme purification of alanine racemase from <u>Pseudomonas sp.</u> ATCC 23,715 through dialysis and enzyme activity assay conditions were similar to those reported by M. Julius, C.A. Free, and G.T. Barry, <u>Methods in Enzymology</u>, XVII A, p. 171-176 (1970).

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